



USE OF SPECIFIC HYBRID PROMOTERS FOR CONTROLLING TISSUE EXPRESSION

FIELD OF THE INVENTION

The present invention relates to the field of biology, and in particular to the field of regulation of expression of genes. It describes new constructs and new vectors that allow a target and high expression of genes. The present invention can be used in many fields, and in particular in the production of recombinant proteins, in the creation of transgenic animal models, in the creation of cell lines, in the development of screening tests, or in gene and cell therapy.

BACKGROUND OF THE INVENTION

The possibility of controlling and directing the expression of genes constitutes a very important stake in the development of biotechnologies. In vitro, it can make it possible to improve the conditions for producing recombinant proteins using particular populations of cells. Still in vitro, it can allow the detection or the revealing of the presence of specific populations of cells in a sample, and also the testing of the properties of a product or the regulation of a gene in a specific population of cells. The control of gene expression is also very important to therapeutic approaches ex vivo or in vivo, in which the possibility of selectively controlling the production of a therapeutic molecule is essential. Indeed, depending on the applications[, depending on] **as well as** the gene to be transferred, it is important to be able to target only certain tissues or certain parts of an organism in order to concentrate the therapeutic effect, and to limit dissemination and side effects.

The targeting of the expression of a given nucleic acid may be achieved according to various approaches. A first approach consists, for example, [in using] **in the use of** transfer agents or vectors [exhibiting] **that exhibit** a given cell specificity. However, the specificity conferred by this type of [vectors] **vector** is generally fairly rough and does not allow targeting of precise populations of cells. Another approach consists [in using] **of the use of** expression signals specific for certain cell types. In this regard, so-called "specific" promoters have been described in the literature, such as [the promoter of the genes encoding] pyruvate kinase **promoter**, the villin **promoter**, GFAP **promoter**, the promoter of the fatty acid-binding intestinal protein, the α -actin promoter of smooth muscle cells, the SM22 promoter, or [alternatively] the promoter [of] **for** the human albumin gene [for example]. However, while these promoters exhibit tissue specificity, they also exhibit[, in return] a relatively low strength. Thus, the great majority of these promoters possess levels of activity [which] **that** are well below [those] **the activity** of so-

called "strong" promoters, generally by a factor of between at least 10 and 100. In addition, it is generally considered that the specificity of a promoter is inversely proportional to its strength, and that the higher the strength **of the promoter**, the higher the level of nonspecific activity.

- 5 [It would therefore be particularly advantageous to be able to have promoters] **In light of the above, it would be advantageous to have a promoter** [which are] **that is** both specific to certain tissues and strong.

SUMMARY OF THE INVENTION

- 10 The subject of the present invention is precisely to provide new constructs [allowing] **that allow** the high and targeted expression of genes. [The invention describes in particular new chimeric promoters allowing] **Provided herein are chimeric promoters that allow** high and specific expression [of the] **of genes in** smooth muscle cells. **The present invention also extends to** [The invention also describes] vectors [containing] **comprising** such promoters, and [their use for
15 the transfer of genes into cells] **the use of such a vector for transferring a gene into a cell *in vitro*, *ex vivo*, and *in vivo*.** The constructs of the **present** invention makes it possible, for the first time, to combine opposing properties, namely high selectivity and a high transcriptional activity. **Thus, the present** [The] invention [thus] offers an efficient means for targeting [of] the expression of genes in smooth muscle cells *in vivo* or *in vitro*, and for the regulation of this
20 expression.

- The **present** invention is based more particularly on the construction of chimeric (or hybrid) promoters comprising regions of different origin and function. More particularly, a first subject of the **present** invention [consists in a hybrid promoter] **extends to a hybrid promoter**
25 comprising:

- all or part of the enhancer region of a strong and ubiquitous promoter/enhancer, and
- a promoter region that allows specific expression in smooth muscle cells.

30

The combination of **an** enhancer [regions] **region** and [of promoters] **a promoter** has already been described in the prior art. Thus, it is known, for example, to couple the CMV enhancer region with nonspecific promoters such as the promoter of the chicken β -actin gene (WO96/13597) in order to increase their strength. However, such [constructs have] **a construct**

has not been described or suggested [with the aim of attempting] **that attempts** to obtain a high expression specific to **a** [the] smooth muscle [cells] **cell**. The **present** invention is partly based on the selection and the combination of particular "enhancer" elements and particular "promoter" elements. [The invention is also based on the demonstration that this combination of elements
5 makes it possible to obtain an expression at high levels.] **In addition, the present invention permits expression at high levels** without affecting the selectivity of the promoter for the target cells of the smooth muscle. **Therefore, the present** [The invention [therefore] provides a particularly advantageous [constructs] **construct** [since they allow] **that allows** targeted production with high levels of molecules in smooth muscle cells. In addition, the present
10 [application] **invention** may also be used both *in vitro* and *in vivo*.

In [the] **a** hybrid [promoters according to] **promoter of the present** invention, the enhancer region and the promoter region are functionally combined[, that is to say such] **so** that the enhancer region exerts a stimulating activity on the activity of the promoter region. Generally,
15 these two regions are therefore genetically linked and sufficiently close to each other to allow the enhancer region to activate the promoter region. Preferably, the distance separating the enhancer region and the promoter region is less than 1 kb, more preferably less than 500 bp. In [the] a particularly preferred [constructs according to the invention] **embodiment**, these two regions are less than 400 bp apart, more preferably less than 200 bp. In addition, as shown in the examples,
20 the respective orientation of these two regions has no significant influence on the activity of **a** [the] hybrid [promoters] **promoter of the present** invention. As a result, the enhancer region may be positioned in the same orientation or in the opposite orientation relative to the direction of the transcription of the promoter region.

25 In a preferred embodiment, the enhancer region is [chosen from] **selected from the group consisting of** the [enhancer region of the] cytomegalovirus immediate-early (CMV-IE) gene **enhancer region**], the enhancer region of the rous sarcoma virus LTR (RSV-LTR), the enhancer region of the SV40 virus, **and** the enhancer region of the EF1 α gene. More preferably, [in the hybrid promoters of the invention,] the enhancer region [is] **of a hybrid promoter of the present**
30 **invention is selected from the group consisting of** the enhancer region of the immediate-early gene of the cytomegalovirus (CMV-IE), and preferably, of the human cytomegalovirus (hCMV-IE). A particular enhancer region consists, for example, of fragment -522 to -63 of the hCMV-IE gene, or [any] **a** fragment [comprising at least part of it and exhibiting] **of the hCMV-IE gene that exhibits** an enhancer activity.

[As regards the promoter region, there is advantageously used for carrying out the invention a region comprising] **Particular examples of a promoter region having applications in a hybrid promoter of the present invention include** all or part of the promoter of the [gene encoding the]
5 α -actin **gene** of smooth muscle cells (SMact) or [of] the SM22 gene. The **promoter** [promoters] of these two genes has been described [for its smooth muscle cell-specific character] **previously** (see in particular Ueyama H. *et al.*, *Mol. Cell Biol.* **4** (1984) 1073-1078; Solway J. *et al.*, *J. Biol. Chem.*, **270** (1995) 13460-13469).

10 A [first particularly advantageous variant] **particular example of a hybrid promoter** of the present invention [consists of a hybrid promoter comprising] **comprises**:

- all or part of the enhancer region of the human cytomegalovirus immediate-early (hCMV-IE) gene, and

15

- all or part of the promoter of the [gene encoding the] α -actin **gene** of smooth muscle cells (Smact), preferably of the human Smact gene.

[A second particularly advantageous variant] **Another example of a hybrid promoter** of the present invention [consists of a hybrid promoter comprising] **comprises**:

20

- all or part of the enhancer region of the human cytomegalovirus immediate-early (hCMV-IE) gene, and

- all or part of the promoter of the SM22 gene, preferably of the mouse SM22 alpha gene.

25

Moreover, in a specific embodiment of the **present** invention, the promoter region is a chimeric region comprising a basal promoter and a sequence conferring tissue specificity, said sequence being derived from the SMact promoter or from the SM22 promoter, or from a combination of both. In this embodiment, the basal promoter may be a "minimal" promoter[, that is to say
30 comprising] **that comprises** only the sequences essential for the transcriptional promoter activity (for example a TATA box). This basal promoter may be the actual basal promoter of the Smact or SM22 gene, or [of] **the actual basal promoter from** a heterologous [origin (β -globin, HSV-TK, SV40, or EF1- α for example)] **gene such as β -globin, HSV-TK, SV40, or EF1- α , for example**. The sequence conferring tissue specificity advantageously comprises part of the

sequence of the Smact promoter (R.T. Shimizu *et al.*, *J. Biol. Chem.* **270** (1995) 7634-7643) and/or of the SM22 promoter (L. Li *et al.* *J. Cell. Biol.* **132** (1996) 849-859; S. Kim *et al.* *J. Clin. Invest.* **100** (1997) 1006-1014; Kemp *et al.*, *Biochem. J.* **310** (1995) 1037-1043).

- 5 A particular [type of] hybrid promoter [according to] **of the present** invention [therefore] comprises:
- all or part of the enhancer region of a strong and ubiquitous promoter/enhancer,
 - a basal promoter, and
 - a sequence conferring tissue specificity comprising all or part of the SMact promoter and/or of
- 10 the SM22 promoter.

For the construction of a hybrid promoter of the present invention, molecular biology techniques known to persons skilled in the art may be used. Thus, the enhancer region and the promoter region (including the basal promoter and the sequence conferring tissue specificity) may be

15 isolated by conventional techniques from nucleic acid libraries or from the total cellular DNA with a technique such as amplification with specific probes. These fragments may also be synthesized artificially using the sequence information available in the prior art. When these fragments are obtained, they can be easily combined [with one another by means of ligases and other restriction enzymes, in order to generate] **together to form a hybrid [promoters] promoter**

20 of the **present** invention **using ligases and restriction enzymes**. In addition, these fragments may be modified by digestion, mutation, insertion or addition of base pairs, either with the aim of facilitating their cloning, or with the aim of modifying their functional properties. Moreover, as indicated above, the fragments may be combined directly with one another or[on the contrary,] separated [by] **from each other with** base pairs [having no] **that have no** significant influence on

25 the activity of the hybrid promoter.

[The] A hybrid [promoters] **promoter** of the **present** invention thus [possess] **possesses** the capacity to express specifically a nucleic acid of interest in [the] smooth muscle cells. The "specific" character of the expression means that the activity of the promoter is significantly

30 much higher in the smooth muscle cells. Although a nonspecific expression may exist in other cells, the corresponding level of activity remains generally very low (negligible) compared with that observed in the smooth muscle cells, generally lower by a factor of at least 10. The results presented in the examples show[in this regard] a difference in expression that may be up to a factor of 140, which reflects the high selectivity of [the promoters] **a hybrid promoter** of the

present invention. [In this regard, the] **The results presented in the examples also show that a hybrid promoter of the present invention exhibits** a high specificity towards smooth muscle cells, since no expression was detected in the endothelial cells [which] **that** exist in the vicinity[.] **or** in the blood vessel, [in particular] **particularly** the artery. The results presented in the

5 examples also show that the strength of [the promoters] **a hybrid promoter of the present invention is much higher than that of [the] a non-hybrid specific [promoters] promoter,** [it being possible for] **with** the difference [to exceed] **in strength exceeding** a factor of 100. These features therefore illustrate the advantageous and unexpected properties of [the] **a hybrid [promoters] promoter of the present invention in terms of strength and specificity[.]** for the

10 expression of **a nucleic [acids] acid** of interest in [the] smooth muscle cells.

[In this regard, another subject of the] **The present invention [relates] further extends** to an expression cassette comprising a nucleic acid [encoding] **that is transcribed into** an RNA of interest, or **encodes** a polypeptide of interest, [placed] **that is** under the control of a hybrid

15 promoter as defined above. Advantageously, [the] **a cassette of the present invention[.] further comprises[, in addition,] a signal [for termination of transcription,] placed 3' of the nucleic acid for termination of transcription.**

Taking into account the cell populations targeted by the cassettes of the **present invention,** the

20 nucleic acid may encode, for example, a protein chosen from:

- [the proteins involved in the cell cycle, such as for example p21 or any other protein inhibiting the cycline-dependent kinases (cdk), the product of the retinoblastoma (Rb) gene, GAX, GAS-1, GAS-3, GAS-6, Gadd 45, Gadd 153, the cyclines A, Band D.] **a protein involved in the cell cycle such as p21 or a protein that inhibits cycline-dependent kinases (cdk), the protein**

25 **product from the expression of the retinoblastoma (Rb) gene, GAX, GAS-1, GAS-3, GAS-6, Gadd 45, Gadd 153, cycline A, cycline B, cycline D;**

-[the proteins inducing apoptosis, such as for example p53, members of the family of apoptosis inducers such as Bas, Bcl-Xs, Bad or any other antagonist of Bcl2 and Bcl-X1.] **a protein that**

30 **induces apoptosis such as p53, Bas, Bcl-Xs, Bad, an antagonist of Bcl2 or an antagonist of Bcl-X1;**

-[the proteins capable of modifying the proliferation of the smooth muscle cells, such as for example an intracellular antibody or an ScFv inhibiting the activity of proteins involved in cell

proliferation, such as for example the Ras protein, map-kinase, or receptors for tyrosine-kinase or for growth factors.] **a protein capable of modifying the proliferation of smooth muscle cells such as an intracellular antibody, an ScFv that inhibits the activity of a protein involved in cell proliferation such as the Ras protein, a MAP-kinase, a receptor for a tyrosine-kinase, or a growth factor;**

-[the proteins for labeling (LacZ, GFP, Luc, secreted alkaline phosphatase (SeAP), growth hormone (GH) and the like) with the aim of carrying out proliferation studies or diagnostic studies.] **a protein for labeling (LacZ, GFP, Luc, secreted alkaline phosphatase (SeAP), growth hormone (GH) and the like) for use in a proliferation study or a diagnostic study;**

- the proteins inducing angiogenesis, such as for example members of the VEGF family, members of the FGF family and more particularly FGF1, FGF2, FGF4, FGFS, angiogenin, EGF, Factor/HGF, members of the angiopoietin family, TGF α , TGF β , TNF α .,

Scatter cytokines and in particular interleukins including IL-1, IL-2, IL-8, angiotensin-2, plasminogen activator (TPA), urokinase (uPA), molecules involved in the synthesis of active lipids (prostaglandins, Cox-I).] **a protein that induces angiogenesis such as a member of the VEGF family or a member of the FGF family (e.g. FGF1, FGF2, FGF4, FGFS, angiogenin, EGF,Factor/HGF), a member of the angiopoietin family, a cytokine such as an interleukin (e.g. IL-1, IL-2, IL-8), angiotensin-2, plasminogen activator (TPA), urokinase (uPA), or a molecule involved in the synthesis of active lipids (e.g. a prostaglandin, Cox-1, etc); or**

-[the transcription factors, such as for example natural or chimeric receptors, comprising a DNA-binding domain, a ligand-binding domain, a transcription activating or inhibiting domain, such as for example the tetR-NLS-VP16 fusion proteins, the fusion proteins derived from the estrogen receptors the fusion proteins derived from the steroid hormone receptors, the fusion proteins derived from the progesterone receptors, the proteins of the CID (chemical inducer of dimerization) system which is described by Rivera et al. (Rivera et al. (1996) *I* A humanized system for pharmacologic control of gene expression, *Nature Medicine*, 2: 1028-1032).] **a transcription factor such as a natural or chimeric receptor that comprises a DNA-binding domain, a ligand-binding domain, a transcription activating or inhibiting domain (e.g. a tetR-NLS-VP16 fusion protein, a fusion protein derived from an estrogen receptor, a fusion protein derived from a steroid hormone receptor, a fusion protein derived from a progesterone receptor), a protein of the CID (chemical inducer of dimerization) system**

described by Rivera et al. (Rivera *et al.* (1996) A humanized system for pharmacologic control of gene expression, *Nature Medicine*, 2: 1028-1032).

It is understood that the present invention is not limited to specific examples of proteins or RNA,
5 but [that it may be used by persons skilled in the art for the] **has applications in the** expression of any nucleic acid in [the] **a** smooth muscle [cells, by] **cell** using simple customary experimental procedures.

[Another subject of the] **The present** invention [relates, in addition to any] **further extends to a**
10 vector comprising a hybrid promoter or a cassette [as defined above] **of the present invention.** [the] **A** vector of the **present** invention may be[, for example,] a plasmid, a cosmid, [or] any DNA not encapsidated by a virus, a phage, an artificial chromosome, a recombinant virus, and the like. It is [preferably] **preferable that a vector of the present invention is a** plasmid or a recombinant virus.

15 [Among the plasmid-type vectors, there may be mentioned all the] **Moreover, all** cloning and/or expression plasmids known to persons skilled in the art and which generally comprise a replication origin **have applications in a vector of the present invention, such as** [There may also be mentioned] new-generation plasmids carrying improved origins and/or markers as
20 described for example in Applications WO96/26270 and PCT/FR96/01414.

[Among the] **Furthermore,** vectors of the recombinant virus type[, there may be preferably mentioned the] **that have applications in the present invention include** recombinant adenoviruses, retroviruses, herpesviruses or adeno-associated viruses. The construction of this
25 type of replication-defective recombinant virus [has] **as well its infectious properties have** been widely described in the literature[, as well as the infectious properties of these vectors] (see in particular S. Baeck and K.L. March (1998), *Circul. Research* vol.82, pp. 295-305), T. Shenk, B.N. Fields, D.M. Knipe, P.M. Howley et al. (1996), *Adenoviridae: the viruses and their replication* (in virology), pp. 211-2148, EDS -Ravenspublishers/Philadelphia, P. Yeh and M.
30 Perricaudet (1997), *FASEB Vol.* 11, pp.615-623).

A recombinant virus that is particularly preferred for carrying out the invention is a defective recombinant adenovirus.

An Adenovirus is linear double-stranded DNA virus of about 36 (kilobases) kb in size. Various serotypes exist whose structures and properties vary somewhat, but which exhibit a comparable genetic organization. More particularly, the recombinant adenoviruses may be of human or animal origin. [As regards the] **With respect to** adenoviruses of human origin, there may be
5 preferably mentioned those classified in group C, in particular the adenoviruses of type 2 (Ad2), 5(Ad5), 7 (Ad7) or 12 (Ad12).

[Among the various] **Preferred** adenoviruses of animal origin[, there may be preferably mentioned the] **include** adenoviruses of canine origin, [and in particular all] **such as** the strains of
10 the CAV2 adenovirus [manhattan or A26/61 strain (ATCC VR-800) for example], **or those disclosed in Application WO94/26914, which is hereby incorporated by reference herein in its entirety.** [Other adenoviruses of animal origin are cited in particular in application WO94/26914 incorporated into the present application by reference.]

15 The genome of the adenovirus comprises [in particular] an inverted repeat sequence (ITR) at each end, an encapsidation sequence (Psi), early genes and late genes. The principal early genes are contained in the E1, E2, E3 and E4 regions. Among these, the genes contained in the E1 region [in particular] are **particularly** necessary for viral propagation. The principal late genes are contained in the L 1 to L5 regions. The genome of the adenovirus Ad5 has been completely
20 sequenced, and is accessible on database (see in particular Genebank M73260). Likewise, portions **of** or even the whole **genome** of other [adenoviral genomes] **adenoviruses** (Ad2, Ad7, Ad12 and the like) have been sequenced.

For their use as recombinant vectors, various constructs derived from adenoviruses have been
25 prepared, incorporating various therapeutic genes. In each of these constructs, the adenovirus was modified [so as] to make it incapable of replicating in the infected cell. Thus, the constructs described in the prior art are adenoviruses [deleted for] **in which** the E1 region, which is essential for viral replication, **has been deleted** at the level at which the heterologous DNA sequences are inserted (Levrero et al., Gene 101 (1991) 195; Gosh-Choudhury et al., Gene 50 (1986) 161).
30 Moreover, in order to improve the properties of the vector, it has been proposed to create other deletions or modifications in the genome of the adenovirus. Thus, a heat-sensitive point mutation was introduced into the ts125 mutant, making it possible to inactivate the 72 kDa DNA-binding protein (DBP) (Van der Vliet et al., 1975). Other vectors comprise a deletion of [another] **the E4 region, which is** essential for replication and/or for viral propagation[, the E4 region]. The E4

region is [indeed] involved in the regulation of the expression of the late genes, in the stability of the late nuclear RNAs, in the extinction of the expression of the host cell proteins, and in the efficiency of the replication of the viral DNA. Adenoviral vectors in which the E1 and E4 regions are deleted possess a transcription background noise and a highly reduced expression of viral
5 genes. Such vectors have been described in applications WO94/28152, WO95/02697, WO96/22378[]]. In addition, vectors carrying a modification at the level of the IVa2 gene have also been described (WO96/10088).

In a preferred embodiment of the invention, the recombinant adenovirus is a group C human
10 adenovirus. More preferably, it is an adenovirus Ad2 or Ad5.

Advantageously, the recombinant adenovirus[used within the framework] of the **present** invention comprises a deletion in the E1 region of its genome, **and in particular a deletion of the E1a and E1b regions**. [Still more particularly, it comprises a deletion for the E1a and E1b
15 regions.] By way of precise example[, there may be mentioned] **are** deletions [affecting] **of** nucleotides 454-3328; 382-3446 or 357-4020 **from the Ad5 genome** [(with reference to the genome of Ad5)].

[According to a preferred variant] **In a preferred embodiment**, [the] **a** recombinant adenovirus
20 [used within the framework of the invention] **of the present invention further** comprises[, in addition,] a deletion in the E4 region of its genome. More particularly, the deletion in the E4 region affects all the open reading frames. There may also be mentioned, by way of precise example, the deletions of nucleotides 33466-35535, or **nucleotides 33093-35535**. Other types of deletions in the E4 region **having applications herein** are described in Applications
25 WO95/02697 and WO96/22378, **which are hereby** incorporated [into the present application[by reference **herein in their entirety**].

[The] **An** expression cassette **of the present invention** may be inserted into various sites of the recombinant genome. [It] **For example**, it may be inserted at the level of the E1, E3 or E4
30 region, as a replacement for deleted nucleotides or as a surplus. It may also be inserted at any other site[.] outside the sequences which are necessary in cis for the production of viruses (ITR sequences and encapsidation sequence).

[The] **A** recombinant [adenoviruses are] **adenovirus of the present invention** is produced in an

encapsidation line, [that is to say] **which** is a line of cells capable of trans-complementing one or more of the functions deficient in the recombinant adenoviral genome. [Among the] **An example of an** encapsidation [lines] **line** known to persons skilled in the art[, there may be mentioned for example] **is** the 293 line into which part of the adenovirus genome has been integrated. More
5 precisely, the 293 line is a human embryonic kidney cell line containing the left end (about 11-12%) of the adenovirus genome serotype 5 (Ad5), comprising the left ITR, the encapsidation region, the E1 region including E1a and E1b, the region encoding the protein pIX and part of the region encoding the protein pIVa2. This line is capable of trans-complementing recombinant adenoviruses [which are defective for] **that lack all or part of** the E1 region, [that is to say
10 lacking all or part of the E1 region,] and of producing viral stocks having high titers. This line is also capable of producing, at permissive temperature (32°C), virus stocks that comprise the heat-sensitive E2 mutation. Other cell lines capable of complementing the E1 region are based upon human lung carcinoma cells A549 (WO94/28152) or on human retinoblasts (Hum. Gen. Ther. (1996) 215). Moreover, lines capable of transcomplementing several functions of the adenovirus
15 have also been described. [In particular, there may also be mentioned] **Particular examples include** lines complementing the E1 and E4 regions (Yeh et al., J. Virol. Vol. 70 (1996) pp 559-565; Cancer Gen. Ther. 2 (1995) 322; Krougliak et al., Hum. Gen. Ther. 6 (1995) 1575), and lines complementing the E1 and E2 regions (WO94/28152, WO95/02697, WO95/27071).

20 Recombinant adenoviruses are usually produced by introducing viral DNA into the encapsidation line, followed by lysis of the cells after about 2 or 3 days (the kinetics of the adenoviral cycle being 24 to 36 hours). For carrying out the method, the viral DNA introduced may be the complete recombinant viral genome, optionally constructed in a bacterium (WO96/25506) or in a yeast (WO95/03400), transfected into the cells. It may also be a recombinant virus used to infect
25 the encapsidation line. The viral DNA may also be introduced in the form of fragments, each containing part of the recombinant viral genome and a region of homology which makes it possible, after introduction into the encapsidation cell, to reconstitute the recombinant viral genome by homologous recombination between the various fragments.

30 After lysis of the cells, the recombinant viral particles are isolated by cesium chloride gradient centrifugation. An alternative method has been described in Patent FR96:08164 [incorporated into the present application], **which is hereby incorporated** by reference **herein in its entirety**.

The **present** invention also [relates] **extends** to a composition [comprising] **that comprises** a

vector [as defined above] **of the present invention** and a chemical or biochemical transfer agent. **As used herein**, [The] **the** term "chemical or biochemical transfer agent" is understood to mean any compound (i.e. other than a recombinant virus) which facilitates the penetration of a nucleic acid into a cell. It may represent cationic nonviral agents such as cationic lipids, peptides,
5 polymers (Polyethylene Imine, Polylysine), nanoparticles; or noncationic nonviral agents such as noncationic liposomes, noncationic nanoparticles or polymers. Such agents are well known to persons skilled in the art.

The invention [also relates] **further extends** to a composition comprising a recombinant virus [as
10 defined above] **of the present invention** and a physiologically acceptable vehicle.

[The] **Moreover, the present** invention [also relates] **extends** to a pharmaceutical composition comprising a vector [as described above] **of the present invention**. [The] **A** pharmaceutical [compositions] **composition** of the **present** invention may be formulated for topical, oral,
15 parenteral, intranasal, intravenous, intramuscular, subcutaneous, intraocular or transdermal **administration** [route and the like].

Preferably, [the] **a** pharmaceutical composition [contains] **of the present invention comprises a** pharmaceutically acceptable [vehicles] **vehicle** for an injectable formulation, in particular for an
20 intravascular injection or injection into smooth muscle tissues. These may be in particular isotonic, sterile, saline solutions (monosodium or disodium phosphate, sodium, potassium, calcium or magnesium chloride and the like, or mixtures of such salts), or dry, [in particular] **e.g.** **a** freeze-dried [compositions] **composition** which, upon addition, depending on the case, of sterilized water or of physiological saline, [allow the constitution of] **forms an** injectable
25 solution. Other excipients [may be used,] such as [for example] a hydrogel **may be used**. [This [hydrogel] **Hydrogel** may be prepared from any biocompatible and noncytotoxic (homo- or hetero-) polymer. Such polymers have [for example] been described in Application WO93/08845. Some of them, [such as in particular] **particularly** those obtained from ethylene and/or propylene oxide, are commercially available. The use of a hydrogel is particularly advantageous for
30 transferring nucleic acids into vascular walls, and in particular into smooth muscle cells of the vascular walls. The doses used for the injection may be adjusted as a function of various parameters, and in particular as a function of the mode of administration used, the aim pursued (labeling, pathology, screening and the like), the gene to be expressed, or alternatively the desired duration of expression. In general, the recombinant viruses according to the invention are

formulated and administered in the form of doses of between 10^4 and 10^{14} pfu, and preferably 10^6 to 10^{10} pfu. The term pfu ("plaque forming unit") corresponds to the infectivity of a viral solution, and is determined by infecting an appropriate cell culture, and measuring the number of plaques of infected cells. The techniques for determining the pfu titer of a viral solution are well

5 documented in the literature. For a use *in vitro* or *ex vitro*, the cassettes, vectors or compositions of the **present** invention may be incubated at conventional doses in the presence of the chosen cell populations. These incubations may be carried out on culture dishes, flasks, fermenters or any other chosen device.

10 Moreover, the **present** invention also [relates] **extends** to a cell modified by a cassette or a vector (in particular an adenovirus) [as described above] **of the present invention**. **As used herein, the term "modified" in describing a** ["Modified"] cell is understood to mean any cell containing a construct [according to the] **of the present** invention. These cells may be used for the production of recombinant proteins *in vitro*. They may also be [intended] **used** for implantation [in] **into** an
15 organism, according to the methodology described in Application WO95/14785. These cells are preferably human smooth muscle cells.

The **present** invention also [relates] **extends** to the use of a hybrid promoter [as defined above] **of the present invention** for the preparation of a composition [intended] for **use in** the expression of
20 a nucleic acid in [the] **a** smooth muscle [cells] **cell** *in vitro*, *ex vivo* or *in vivo*.

In addition, the present invention extends [The invention also relates] to the use of a hybrid promoter [as defined above] **of the present invention** for the preparation of a composition intended for the expression of a nucleic acid in [the] **a** smooth muscle [cells] **cell** *in vivo* and not
25 in [the] **an** endothelial [cells which exist]] **cell that exists** in the vicinity in the artery **in which the smooth muscle cell is located**.

As a result of their smooth muscle cell-specific character, the constructs according to the invention can also be used for the creation of animal models for vascular pathologies, [or] for
30 carrying out labeling studies, or in methods for [the detection of or] screening for the presence of **a** smooth muscle [cells] **cell** in a sample.

[The subject of the] **The present invention** [is] also **extends to** a method of producing **a** recombinant [proteins comprising introduction into a cell population of a vector as defined above.

the culture of said recombinant cell population, and the recovery of said protein produced]
**protein that comprises the steps of introducing a vector of the present invention into a cell
population to form a recombinant cell population, culturing said recombinant cell
population, and recovering the protein produced.** Advantageously, smooth muscle cells are
5 used for carrying out the method according to the invention. These may be established lines or
primary cultures.

The present application will be described in greater detail with the aid of the following examples
which should be considered as illustrative and nonlimiting.

10

LEGEND TO THE FIGURES

Table I: Relative activities of the hybrid promoters (hSM α -actin) evaluated in transient
transfections *in vitro* into rabbit smooth muscle cells in primary culture (rabbit SMC), into
ECV304 cells, into myoblasts C2C12, into HeLa cells, into NIH 3T3 cells, into carcinoma cells
15 TU182, as well as into the renal cells 293. The relative activity of each promoter is expressed as a
percentage of the luciferase activity obtained with the plasmid pCMV-leadTK. Enh-X: the
enhancer sequence of hCMV-IE is cloned upstream of the X promoter in its normal orientation.
HnE-X: the enhancer sequence of hCMV-IE is cloned upstream of the X promoter in the opposite
orientation.

20

Table II: Relative activities of the hybrid promoters (mSM22) evaluated in transient transfections
in vitro into rabbit smooth muscle cells in primary culture (rabbit SMC), into ECV304 cells, into
myoblasts C2C12, into HeLa cells, into NIH 3T3 cells, into carcinoma cells TU182, as well as
into the renal cells 293. The relative activity of each promoter is expressed as a percentage of the
25 luciferase activity obtained with the plasmid pCMV-leadTK. Enh-X: the enhancer sequence of
hCMV-IE IS cloned upstream of the X promoter in its normal orientation. HnE-X: the enhancer
sequence of hCMV-IE is cloned upstream of the X promoter in the opposite orientation.

Figure 1: Schematic representations of the plasmids whose expression cassette contains the
30 hSM α -actin hybrid promoter.

Figure 2: Schematic representations of the plasmids whose expression cassette contains the
hybrid promoter mSM22 α .

Figure 3: Activities of the hybrid promoters evaluated in transient transfections *in vitro* into rabbit smooth muscle cells in primary culture (rabbit SMC), into endothelial cells derived from a human umbilical cord carcinoma (ECV304), into mouse myoblasts (C2C12) as well as into epithelial cells derived from a human cervical carcinoma (HeLa). The relative activity of each promoter is expressed as a percentage of the luciferase activity obtained with the plasmid pCMV-leadTK. Enh-X: the enhancer sequence of hCMV-IE is cloned upstream of the X promoter in its normal orientation. hnE-X: the enhancer sequence of hCMV-IE is cloned upstream of the X promoter in the opposite orientation.

Figure 4: Activities of the hybrid promoters evaluated in transient transfections *in vitro* into mouse embryonic fibroblasts (NIH 3T3), into cells derived from a human ENT carcinoma (TU182), as well as into transformed human embryonic renal cells (293). The relative activity of each promoter is expressed as a percentage of the luciferase activity obtained with the plasmid pCMV-leadTK. Enh-X: the enhancer sequence of hCMV-IE is cloned upstream of the X promoter in its normal orientation. hnE-X: the enhancer sequence of hCMV-IE is cloned upstream of the X promoter in the opposite orientation.

Figure 5: Activities of the hybrid promoters evaluated in gene transfer *in vivo* into the C57BL6 mouse cranial tibial muscle. The relative activity of each promoter is expressed as a percentage of the luciferase activity obtained with the plasmid pCMV-leadTK.

MATERIALS AND METHODS

The methods conventionally used in molecular biology such as preparative extractions of plasmid DNA, centrifugation of plasmid DNA in cesium chloride gradient, agarose gel electrophoresis, purification of DNA fragments by electroelution, precipitation of plasmid DNA in saline medium with ethanol or isopropanol, or transformation in *Escherichia coli* are well known to persons skilled in the art, and are abundantly described in the literature (Sambrook et al. "Molecular Cloning, a Laboratory Manual", Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., 1989).

The plasmid pGL3-Basic, used for the cloning of the various promoter regions, is of commercial origin (Promega Corporation). The plasmids pCMV13 (Clontech Laboratories Inc.) and pUC18 (Boehringer Mannheim) are also of commercial origin.

The enzymatic amplification of DNA fragments by the PCR technique (Polymerase Chain Reaction) may be carried out using a DNA Thermal Cycler (Perkin Elmer Cetus) according to the manufacturer's recommendations.

- 5 The electroporation of plasmid DNA into *Escherichia coli* cells may be carried out with the aid of an electro orator (Bio-Rad) according to the manufacturer's recommendations.

The verification of the nucleotide sequences may be carried out by the method developed by Sanger et al. (*Proc. Natl. Acad. Sci. USA*, **74** (1977) 5463-5467) using the kit distributed by

- 10 Applied Bios stems according to the manufacturer's recommendations.

EXAMPLES

EXAMPLE 1: Construction of hybrid promoters and of expression plasmids containing them

1.1. Hybrid promoters hSM α -actin

- 15 Plasmid phSMact: The high-molecular-weight genomic DNA was prepared according to the method described by Sambrook et al. ("Molecular Cloning, a Laboratory Manual", Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., 1989) from a primary culture of human aortic smooth muscle cells (Clonetics).

This DNA was used as template for a first amplification by PCR using the following primers:

20

-Primer 6417 (5' GA TGGTCCCT ACTT A TGCTGCT A 3') (SEQ ID NO:1) starting at position -1034 (promoter region) of the human specific smooth muscle α -actin gene (Ueyama H. et al., *Mol. Cell. Biol.*, **4** (1984) 1073-1078. Access Genbank D00618);

- 25 -Primer 6418 (5' CTTCCATCATACCAAACACTACATA 3') (SEQ ID NO:2) at position 1974 of the D00618 sequence situated inside the first intron of the hSMact gene.

The reaction mixture comprises 1 mg of genomic DNA, 10 pmol of each of the two primers (6417 and 6418), 100 mM of each deoxyribonucleotide (dATP, dCTP, dGTP, dTTP), 2 mM

- 30 MgCl₂ and 5 units of Taq DNA polymerase (Perkin Elmer). The reaction volume is brought to 50 ml adjusted to the optimum PCR buffer concentration recommended by Perkin Elmer.

The PCR amplification is carried out in MicoampTM tubes (Perkin Elmer) with [theaid] the aid of a thermocycler PTC-100 (MJ Research, Inc.). This amplification consists of a denaturing step at

95°C for 2 min followed by 30 cycles comprising a denaturing step of 15 sec at 95°C, an annealing step of 30 sec at 60°C and an extension step of 1 min at 72°C. These thirty cycles are followed by an additional extension of 5 min and then the PCR reactions are stored at 10°C.

One microliter of this reaction was collected from this first reaction and then diluted in 10 ml of water. Next, 1 ml of this dilution was used to carry out a second PCR under the same conditions as the first (above) but with a different pair of primers:

-Primer 6453 (5' CTGCTAAATTGctcgagGACAAATTAGACAAA 3') (SEQ ID NO:3), this primer introduces a XhoI site (underlined lower-case letters) upstream of the hSMact promoter (position -680).

-Primer 6456 (5' CCCTGACAAAagcttGGCTGGGCTGCTCCACTGG 3') (SEQ ID NO:4), this primer introduces a HindIII site at position +30 of hSMact.

After analysis on an agarose gel and then purification, the DNA fragment amplified by PCR is digested for 3 hours at 37°C with XhoI and HindIII and then cloned into the vector pGL3-Basic (Promega) previously digested with these same restriction enzymes[, in order] to generate the plasmid phSMact (Figure 1).

Plasmids pXL3130 and pXL3131: A DNA fragment corresponding to the enhancer region of the promoter of the human cytomegalovirus IE (hCMV-IE) gene between positions -522 and -63 relative to the site of initiation of transcription was amplified by PCR using the plasmid pCMV β as a template and the oligonucleotides 8557 (5' ATC GAC GCG TGC CCG TTA CAT MC TTA CGG 3') (SEQ ID NO: 5) and 8558 (5' ATC GAC GCG TCC GCT CGA GCG TCA ATG GGG CGG AGT TG 3') (SEQ ID NO: 6) as primers. This fragment was digested with MluI and was then cloned into the plasmid phSMact previously digested with MluI, and then treated with alkaline phosphatase. Depending on the direction of insertion of the fragment, two different plasmids were obtained: pXL3130 and pXL3131. The schematic representations of these plasmids are set forth in [the figure (Figure 1) **Figure 1**. These plasmids contain, in the form of an MluI-NcoI fragment,] a hybrid promoter [consisting of the enhancer of the promoter of] in the form of an **MluI-NcoI fragment that consists of the enhancer of the promoter** for the hCMV-IE gene and of the promoter of the hSM α -actin gene.

1.2. Hybrid promoters mSM22

Plasmid pmSM22: The high-molecular-weight genomic DNA was prepared according to the method described by Sambrook et al. ("Molecular Cloning, a Laboratory Manual", Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., 1989) from a Balbc mouse liver.

This DNA was used as template for a PCR amplification using the following primers:

5

-Primer 6517:

(5' CCAGGCTGCActcgagACTAGTCCCCACCMCTCGA 3') (SEQ ID NO:7) this primer introduces an XhoI site (underlined lower-case letters) at position -436 of the promoter of the mouse SM22 alpha gene (Solway J. et al., *J. Biol. Chem.*, **270** (1995) 13460-13469; Access
10 Genbank L41161).

-Primer 6518:

(5' TCGTTTGaagctGGAAGGAGAGTAGCTTCGGTGTC 3') (SEQ ID NO:8), this primer introduces a HindIII site at position +43 of mSM22 alpha.

15 A reaction mixture comprising 1 mg of mouse genomic DNA, and 10 pmol of each of the two primers (6517 and 6518) was prepared with the same reagents as for hSMact and at the same concentrations, followed by a PCR amplification carried out under the same conditions (see Example 1.1.).

20 After analysis on an agarose gel and then purification, the DNA fragment amplified by PCR was digested for 3 hours at 37°C with XhoI and HindIII, and then cloned into the vector pGL3-Basic (Promega) **that was** previously digested with these same restriction enzymes. The resulting plasmid was designated pmSM22 (Figure 2).

25 Plasmids pXL3152 and pXL3153: A DNA fragment corresponding to the enhancer region of the promoter of the hCMV-IE gene between positions -522 and -63 relative to the site of initiation of transcription was amplified by PCR using the plasmid pCMVr3 as template and the oligonucleotides 8557

(5' ATC GAC GCG TGC CCG TTA CAT AAC TTA CGG 3') (SEQ ID NO:5) and 8558 (5' ATC
30 GAC GCG TCC GCT CGA GCG TCA A TG GGG CGG AGT TG 3') (SEQ ID NO:6) as primers. This fragment was digested with MluI and was then cloned into the plasmid pmSM22 previously digested with MluI and treated with alkaline phosphatase. Depending on the direction of insertion of the fragment, two different plasmids were obtained: pXL3152 and pXL3153. The schematic representations of these plasmids are [assembled] **set forth** in Figure 2. These plasmids

contain, in the form of an MluI-NcoI fragment, a hybrid promoter consisting of the enhancer of the promoter of the hCMV-IE gene and of the promoter of the mSM22 gene.

1.3. Control plasmid pCMV-leadTK

- 5 The expression vector pCGN previously described by Tanaka et al. (*Cell*, 60 (1990) 375-386) contains the CMV promoter (-522/+72) fused with the "leader" of the HSV tk gene (+51/+101) upstream of a sequence encoding the hemagglutinin epitope. The plasmid pCGN (10 ng) was used as template for PCR amplification. The PCR reaction as well as the amplification were carried out under the same conditions as those used for hSMact and mSM22 (Examples 1.1 and 10 1.2). The primers which were used are the following:

-Primer 6718

(5' CCCGTTACATAACTTACGGTAAATGGCCCG 3') (SEQ ID NO: 9), this primer hybridizes with the CMV promoter at position -522 (8 nucleotides downstream of the *EcoRI* site of pCGN).

15

-Primer 6719

(5' **g**GACGCGCTTCTACAAGGCGCTGGCCGAA 3') (SEQ ID NO:10), this primer hybridizes up to position 101 of the tk "leader". The first **nucleotide** [nucleotides] G in bold is intended to restore the NcoI site of pGL3-Basic as will be clearly stated below.

20

The PCR fragment thus obtained is purified and then phosphorylated with the aid of T4 phage polynucleotide kinase (New England Biolabs). In parallel, the vector pGL3-Basic (Promega) was linearized with NcoI, purified and then treated with Klenow DNA polymerase (Boehringer Mannheim) in order to fill the NcoI site. This vector is then dephosphorylated with the aid of 25 alkaline phosphatase (Boehringer Mannheim) and then used for the insertion of the phosphorylated PCR fragment. Thus, the guanosine (G) of the primer 6719 makes it possible to restore the NcoI site only when the CMV-tk leader fragment is oriented with the 5' part (primer 6718, position -522 of CMV) downstream of the HindIII site of pGL3-Basic and its 3' end (primer 6719, tk leader) is ligated into the NcoI site of pGL3-Basic (first ATG of luciferase). The 30 plasmid thus obtained is designated pCMV-leadTK.

EXAMPLE 2: Specificity of the hybrid promoters *in vitro*

This example illustrates the tissue specificity properties of a hybrid promoter of the present invention *in vitro*.

2.1. Cell cultures

The rabbit smooth muscle cells (SMC) are cultured in DMEMTM medium (Life Technologies Inc.) supplemented with 20% fetal calf serum (FCS). The ECV304 cells are cultured in 199TM medium (Life Technologies Inc.) supplemented with 10% FCS. C2C12 myoblasts, HeLa cells, NIH 3T3 cells, and TU182 cells are cultured in DMEMTM medium supplemented with 10% FCS. The 293 cells are cultured in MEMTM medium (Life Technologies Inc.) supplemented with pyruvate, nonessential amino acids and 10% FCS. All the cultures are carried out in an incubator at 37°C in a humid atmosphere and under a CO₂ partial pressure of 5%.

2.2. Transfections *in vitro*

The transfections are carried out in 24-well plates, and each transfection is carried out three times. Twenty-four hours before transfection, the cells are inoculated: (i) at 5×10^4 cells per well for the rabbit smooth muscle cells, the ECV304, NIH 3T3 and HeLa cells. (ii) at 10^5 cells per well for the TU182 cells, (iii) at 3×10^4 cells per well for the C2C12 cells, and (iv) at 2×10^5 cells per well for the 293 cells.

For each well, 500 ng of plasmid DNA (250 ng of plasmid of interest and 250 ng of pUC18) are mixed with the cationic lipid RPR120535 B (WO 97/18185) in an amount of 6 nmol of lipid per μg of DNA in DMEMTM medium (20 μ final) comprising 150 mM of NaCl and 50 mM of bicarbonate. After 20 minutes at room temperature, the 20 μl DNA/lipid mixture are brought into contact with the cells[, in the absence of FCS.] for 2 hours. The culture medium is then supplemented with FCS to obtain the percentage of FCS required for the culture of each cell type.

Forty-eight hours after the transfection, the culture medium is removed and the cells are rinsed twice with PBS (Life Technologies Inc.). The luciferase activity is then determined with the aid of the Luciferase Assay SystemTM kit (Promega Corporation) according to the manufacturer's recommendations.

2.3. Specific activities of the hybrid promoters

The relative luciferase activities of the hybrid promoters (relative to the pCMV-lead TK promoter) measured *in vitro* in seven different cell types are [assembled] **set forth** in Figures 3 and 4, as well as in Tables I and II. These results show that for the promoters hSMact (Table I) and mSM22 (Table II), the relative activity is a value that clearly reflects the specificity of these promoters for smooth muscle cells; a specificity which has already been described in the literature (Skalli et al., *J. Histochem. Cytochem.*, 37 (1989) 315-321; Shimizu et al., *J. Biol. Chem.*, 270

(1995) 7631-7643; Li et al., *J. Cell Biol.*, 132 (1996) 849-859). Indeed, the relative activity in rabbit SMCs is at least 5 times higher than those observed in the other cell types: (i) from 5 to 20 times higher for the promoter hSMact (Table I), and (ii) from 5 to 25 times higher for the promoter mSM22 (Table II).

5

The results presented in Tables I and II show clearly that, in the smooth muscle cells, the activity of the four hybrid promoters according to the invention (Enh-hSMact, hnE-hSMact, Enh-mSM22 and hnE-mSM22) are comparable, in terms of strength, to that of the CMV promoter. Moreover, the relative activity of each of these promoters in another cell type is at least 10 times lower than that of the hSMact hybrid promoters, and at least 4 times lower for the mSM22 hybrid promoters than that observed in the smooth muscle cells: (i) from 10 to 140 times for the hSMact hybrid promoters, and (ii) from 4 to 55 times for the mSM22 hybrid promoters. These hybrid promoters therefore preserve the same tissue specificity as that observed for the specific promoters hSMact and mSM22.

15

These results show, in addition, that the orientation of the enhancer region in the hybrid promoters of the invention has no significant influence on their activity.

The four hybrid promoters of the present invention therefore possess *in vitro* activity in smooth muscle cells that is as high as that for the CMV promoter (which is reputed to be a strong promoter), while preserving a tissue specificity which is comparable to, or even greater than that of the hSMact and mSM22 promoters.

EXAMPLE 3: Specificity of the hybrid promoters *in vivo*

This example illustrates the tissue specificity properties of the hybrid promoters of the invention *in vivo*.

25

3.1. Gene transfer into the skeletal muscle

The various plasmids were injected, intramuscularly, into the cranial tibial muscle of 5-week old female C57BL6 mice. Each plasmid, diluted in a solution of NaCl at 150 mM final, is injected in an amount of 10 μ g per muscle. Three days after injection, the muscles are collected in 2 ml of Cell Culture Lysis ReagentTM buffer (Promega Corporation), and ground with the aid of a Diox homogenizer (Heidolph). The ground product is then centrifuged for 15 minutes at 4000 g, and then the luciferase activity is evaluated with the aid of the Luciferase Assay SystemTM kit (Promega Corporation) according to the manufacturer's recommendations.

30

3.2. Activities of the hybrid promoters in the skeletal muscle *in vivo*

The relative activities of the two specific promoters (hSMact and mSM22) as well as those of two of the hybrid promoters of the invention (Enh-hSMact and Enh-mSM22) were also evaluated *in vivo* after transfer of naked DNA into the mouse cranial tibial muscle. The results assembled in Figure 5 show that the activity of the Enh-hSMact promoter is 100 times lower than that of the CMV promoter. Likewise, the activity of the Enh-mSM22 promoter is 17 times lower than that of the CMV promoter. Thus, the tissue specificity observed *in vitro*, and in particular in the C2C12 cells which constitute the model which is closest to that used *in vivo*, is therefore conserved *in vivo*.

EXAMPLE 4: Construction of recombinant adenoviruses expressing the GAX protein under the control of specific hybrid promoters

This example is intended to describe an adenoviral vector carrying the gene encoding the GAX protein operably linked to the hybrid promoter of the invention composed of the CMV enhancer and the SM α -actin promoter (enh-hSMact).

The human *gax* gene contains 912 base pairs and encodes a transcription factor of 303 amino acids which is involved in the arrest of cell growth (growth-arrest-specific homeobox) and which has a role on the proliferation of the human smooth muscle cells. This homeodomain-containing gene was initially isolated from the aorta and is expressed in particular in adult cardiovascular tissues (Gorski et al. 1993).

The sequence of the human *gax* gene has been [to clone] **cloned** from a skeletal muscle cDNA library by PCR (Polymerase Chain Reaction) using[, as primer,] a sequence derived from the human *gax* gene and published by Walsh *et al.* (Genomics (1994), 24, p535) **as a primer**. The sequence was then cloned into the expression vector pXL3297. This plasmid is derived from the plasmid Bluescript (Stratagene) containing the human CMV IE enhancer/promoter (-522/+72) (Cell (1985), 41, p521) and the SV 40 poly A (2538-2759) (GenBank SV4CG locus). [The construction uses] **Also**, the plasmid pXL3130 described in Example 1 (Figure 1), into which the SM α -actin promoter had been previously introduced, **was used in its construction**. The plasmid pXL3297 is an expression vector containing the human *gax* gene. It was digested with the enzymes HindIII and AvrII in order to introduce the human *gax* gene into the plasmid pXL3282, which had also been digested with the enzymes HindIII and AvrII in order to produce plasmid pXL 3300. As indicated in Figure 6, in which the various steps of the construction of the plasmids are described in detail, the final plasmid pXL3310, contains an expression cassette consisting of

the CMV-IE enhancer and the SM α -actin promoter (pSMA), combined according to the invention and operably linked to the gene encoding the human GAX protein as well as the SV40 poly A termination signal.

- 5 The human gax gene expression cassette is then introduced into a recombinant human adenovirus of serotype 5 (AdS) [deleted for] **in which** the E1 and E3 regions **had been deleted** by co-transfection **in encapsidation cells**, and homologous recombination between the plasmid carrying the cassette for expressing the gax gene and the adenovirus[, in encapsidation cells]. These cells are preferably the 293 line. The production of an adenovirus stock containing the cassette for
- 10 expressing the human gax gene results from the lysis of the encapsidation cells 2 or 3 days after infection and isolation of the recombinant viral particles with cesium chloride gradient centrifugation.

- The viral particles are then used to study the expression of the human gax gene under the control
- 15 of the promoter of the invention in smooth muscle cells.

The expression of the GAX protein is checked 24 hours after infection of the primary smooth muscle cells with immunofluorescence or western blotting using the rabbit anti-gax polyclonal antibodies.

20

The expression of the messenger RNAs is analyzed 24 hours after the infection of the smooth muscle cells with dot blotting or northern blotting using an oligonucleotide whose sequence is present in the gax gene.

- 25 The analysis of the biological activity of the adenovirus encoding the gax gene is carried out in the following manner:

- Smooth muscle cells at the exponential growth phase are infected with an adenovirus containing the gene encoding the GAX protein under the control of the enh-hSMact promoter in the absence
- 30 and in the presence of 125 ng of lipofectin (48-well plates). The adenoviruses (in variable dilutions) and the lipofectamine are incubated for 30 minutes at room temperature in a serum-free medium. The mixture or the virus alone is brought into contact with the cells for one hour at 37°C. At the end of the period of infection, the medium containing the virus is removed and the cells are incubated in DMEM medium containing 0.5% SVF. Within the 24 hours following the

infection, the culture medium is replaced with a growth medium for half of the cultures and the incubation is continued for 48 hours to allow the cells to enter the S phase. For the other half of the cultures, a weakly mitogenic medium is added in order to maintain the cells in quiescence. The viable cells are counted 72 hours after the infection using the Alamar protocol.

TABLE 1

5

Plasmids
(Promoters)

	Without Plasmid	PCMV-leadTK	PhSMact	pXL3130	pXL3131
	(-)	(CMV-leadTK)	(hSMact)	(Enh-hSMact)	(hnE-hSMact)
Rabbit SMC	< 0.01	100.00	0.67 ± 0.06	107.10 ± 11.23	77.9 ± 7.93
ECV304	< 0.01	100.00	0.12 ± 0.05	5.19 ± 0.99	4.68 ± 0.77
C2C12	< 0.01	100.00	0.11 ± 0.01	4.48 ± 0.28	2.81 ± 0.09
HeLa	< 0.01	100.00	0.03 ± 0.01	10.31 ± 1.13	8.00 ± 0.19
NIH 3T3	< 0.01	100.00	0.08 ± 0.01	4.25 ± 0.08	3.66 ± 0.28
TU182	< 0.01	100.00	0.03 ± 0.00	6.15 ± 0.47	8.46 ± 0.41
293	< 0.01	100.00	0.04 ± 0.00	0.77 ± 0.07	0.72 ± 0.07

10

TABLE 2

Plasmids
(Promoters)

	Without Plasmid	PCMV-leadTK	PmSM22	pXL3152	pXL3153
	(-)	(CMV-leadTK)	(mSM22)	(Enh-mSM22)	(hnE-hSM22)
Rabbit SMC	< 0.01	100.00	2.13 ± 0.29	94.61 ± 13.43	100.21 ± 23.20
ECV304	< 0.01	100.00	0.42 ± 0.13	13.81 ± 0.35	11.83 ± 1.95
C2C12	< 0.01	100.00	0.20 ± 0.01	11.19 ± 1.84	8.37 ± 0.08
HeLa	< 0.01	100.00	0.04 ± 0.01	12.87 ± 1.04	15.90 ± 1.92
NIH 3T3	< 0.01	100.00	0.32 ± 0.01	9.49 ± 0.41	9.59 ± 0.53
TU182	< 0.01	100.00	0.08 ± 0.01	25.78 ± 3.04	26.37 ± 0.98
293	< 0.01	100.00	0.09 ± 0.00	2.29 ± 0.02	1.78 ± 0.03

15

CLAIMS

1. Hybrid promoter comprising:
 - all or part of the enhancer region of a strong and ubiquitous promoter/enhancer, and
 - 5 -a promoter region allowing specific expression in the smooth muscle cells.
- 10 2. Hybrid promoter according to claim 1, characterized in that the enhancer region is chosen from the enhancer region of the cytomegalovirus immediate- early (CMV-IE) gene, the enhancer region of the rous sarcoma virus L TR (RSV-L TR), the enhancer region of the SV40 virus, and the enhancer region of the EF1a gene.
3. Hybrid promoter according to claim 2, characterized in that the enhancer region is the enhancer
- 15 region of the immediate-early gene of the cytomegalovirus (CMV-IE), preferably of the human cytomegalovirus (hCMV-IE).
4. Hybrid promoter according to claim 1, characterized in that the promoter region comprises all or part of the promoter of the gene encoding the α -actin of smooth muscle cells (SMact) or of the SM22 gene.
- 20 5. Hybrid promoter comprising:
 - all or part of the enhancer region of the human cytomegalovirus immediate-early (hCMV-IE) gene, and
 - all or part of the promoter of the gene encoding the α -actin of smooth muscle cells (SMact).
6. Hybrid promoter comprising:
- 25 -all or part of the enhancer region of the human cytomegalovirus immediate-early (hCMV-IE) gene, and
- all or part of the promoter of the SM22 gene.
7. Hybrid promoter according to claim 1, characterized in that the promoter region comprises a basal promoter and a sequence conferring tissue specificity, said sequence being derived from the
- 30 SMact promoter and/or from the SM22 promoter.
8. Expression cassette comprising a nucleic acid encoding an RNA or a polypeptide of interest, placed under the control of a hybrid promoter according to one of claims 1 to 7.

9. Cassette according to claim 8, characterized in that it comprises in addition, a signal for termination of transcription.
10. Cassette according to claim 8 or 9, characterized in that the nucleic acid encodes a protein
5 chosen from the proteins involved in the cell cycle, the proteins inducing apoptosis, the proteins capable of modifying the proliferation of the smooth muscle cells, the proteins inducing angiogenesis and the transcription factors.
11. Vector comprising a hybrid promoter according to claim 1 or a cassette according to claim 8.
- 10 12. Vector according to claim 11, characterized in that it is a plasmid, a cosmid or any DNA not encapsidated by a virus.
13. Vector according to claim 11, characterized in that it is a recombinant virus, preferably derived from an adenovirus, a retrovirus, a herpesvirus or an adeno-associated virus.
- 15 14. Composition comprising a vector according to claim 12 and a chemical or biochemical transfer agent.
15. Composition comprising a recombinant virus according to claim 13 and a physiologically acceptable vehicle.
16. Cell modified by a cassette according to claim 8 or a vector according to claim 11.
- 20 17. Use of a hybrid promoter according to one of claims 1 to 7 for the preparation of a composition intended for the selective expression of a nucleic acid in the smooth muscle cells.
18. Use of a hybrid promoter according to one of claims 1 to 7 for the preparation of a composition intended for the expression of a nucleic acid in the smooth muscle cells and not in the endothelial cells which are found in the vicinity of the blood vessel.

25

ABSTRACT OF THE DISCLOSURE

The invention concerns a hybrid promoter comprising all or part of the enhancer region of a strong and ubiquitous promoter/enhancer; a promoter region enabling specific expression in smooth muscle cells, and all vector or cell containing said promoter, and their uses.